

Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity¹⁻³.

General eDNA Assay Information

Target Species: <i>Heteromastus filobranchus</i>	eDNA qPCR Tool: eHEFI7	Gene Target: MT-CYB
Species Code: an-HEFI	eDNA qPCR Format: Taqman	Published in:

eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD <u>0.7</u>	95% CI <u>0.5-1.1</u>	Copies	LOQ <u>2.8</u>	95% CI <u>2.1-4.2</u>	Copies	LOB <u>0</u>	hits/8
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LOQ_{continuous} 20 Copies/Rxn

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code⁴. When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

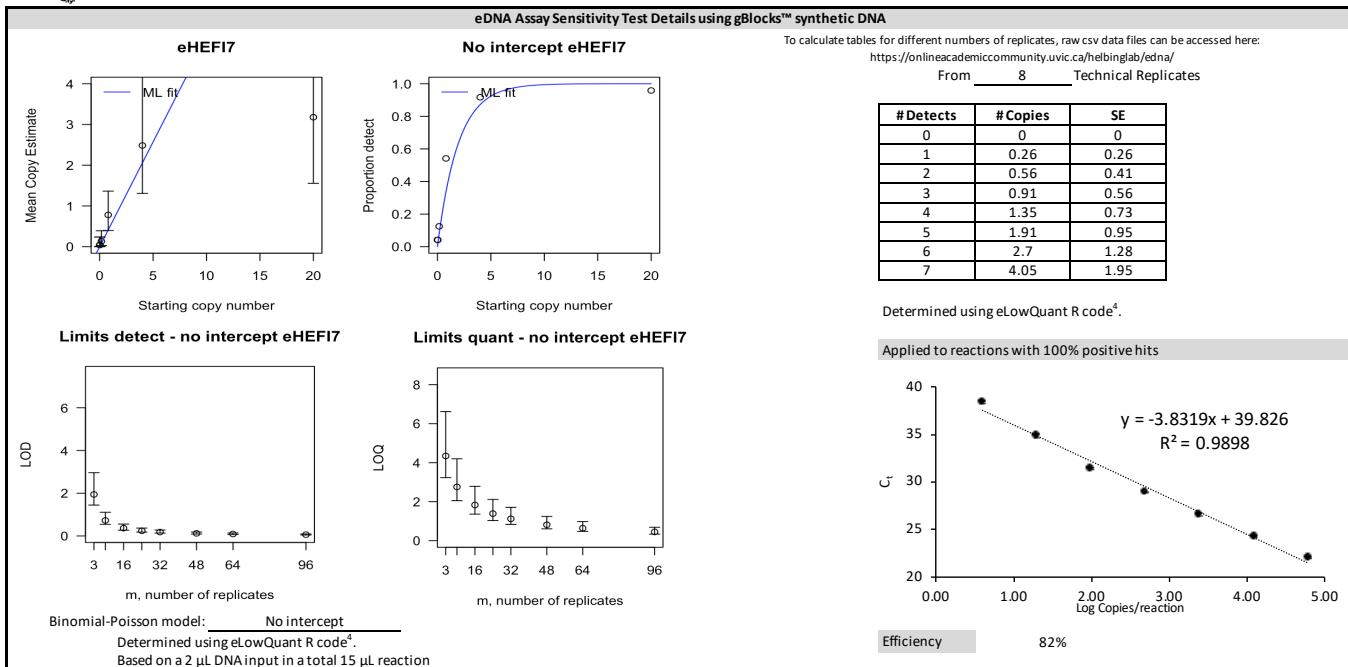
eDNA Assay Specificity Test Information

Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Species	Common Name (Species)	Detection	Specimens	# Voucher
				Sample Sources/Locations
an-CACA	<i>Capitella capitata</i>	No	3	British Columbia
an-DEGR	<i>Decamastus gracilis</i>	No	4	British Columbia
an-GLNA	<i>Glycera nana</i>	No	2	British Columbia
an-HEFI	<i>Heteromastus filobranchus</i>	Yes	4	British Columbia
an-NOHE	<i>Notomastus hemipodus</i>	No	2	British Columbia
an-PRJU	<i>Prionospio (Prionospio) jubata</i>	No	2	British Columbia
an-PRLI	<i>Prionospio (Minuspio) lighti</i>	No	4	British Columbia
an-PRMU	<i>Prionospio (Minuspio) multibranchiata</i>	No	1	British Columbia
an-RIP1	Hydrothermal vent worm (<i>Ridgeia piscesae</i>)	No	3	British Columbia
ma-CALUfa	Domestic dog (<i>Canis lupus familiaris</i>)	No	1	British Columbia
ma-FECA	Domestic cat (<i>Felis catus</i>)	No	1	British Columbia
ma-HOSA	Human (<i>Homo sapiens</i>)	No	1	Netherlands

References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA. 2020; 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 3: 970-981. doi: 10.1002/edn3.220



Field Sample Validation				
Sample Type	Known Presence	# Samples	Detected	Location
Water	Y	280	Y	Vancouver, BC, Canada

Abbreviations					
95% CI	95% Confidence interval	LOQ	Limit of quantification		
eDNA	Environmental DNA	MT-CYB	Mitochondrial cytochrome b gene		
gDNA	Total genomic DNA extracted from voucher specimen	NTC	qPCR no template control		
LOB	Limit of blank	qPCR	Quantitative real-time polymerase chain reaction		
LOD	Limit of detection	SE	Standard error		