



### **Helbing/Langlois Laboratory eDNA Technical Bulletin**

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity<sup>1-3</sup>.

#### **General eDNA Assay Information**

Target Species: Southern bog lemming (*Synaptomys cooperi*)  
Species Code: ma-SYCO

eDNA qPCR Tool: eSYCO1  
eDNA qPCR Format: TaqMan

Gene Target: MT-ND2  
Published in:

#### **eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA**

LOD	0.2	95% CI	0.2-0.4	Copies/Rxn	LOQ	0.9	95% CI	0.6-1.6	Copies/Rxn	LOB	0	hits/8
				LOQ <sub>continuous</sub>	4			Copies/Rxn				

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code<sup>4</sup>.

When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

#### **eDNA Assay Specificity Test Information**

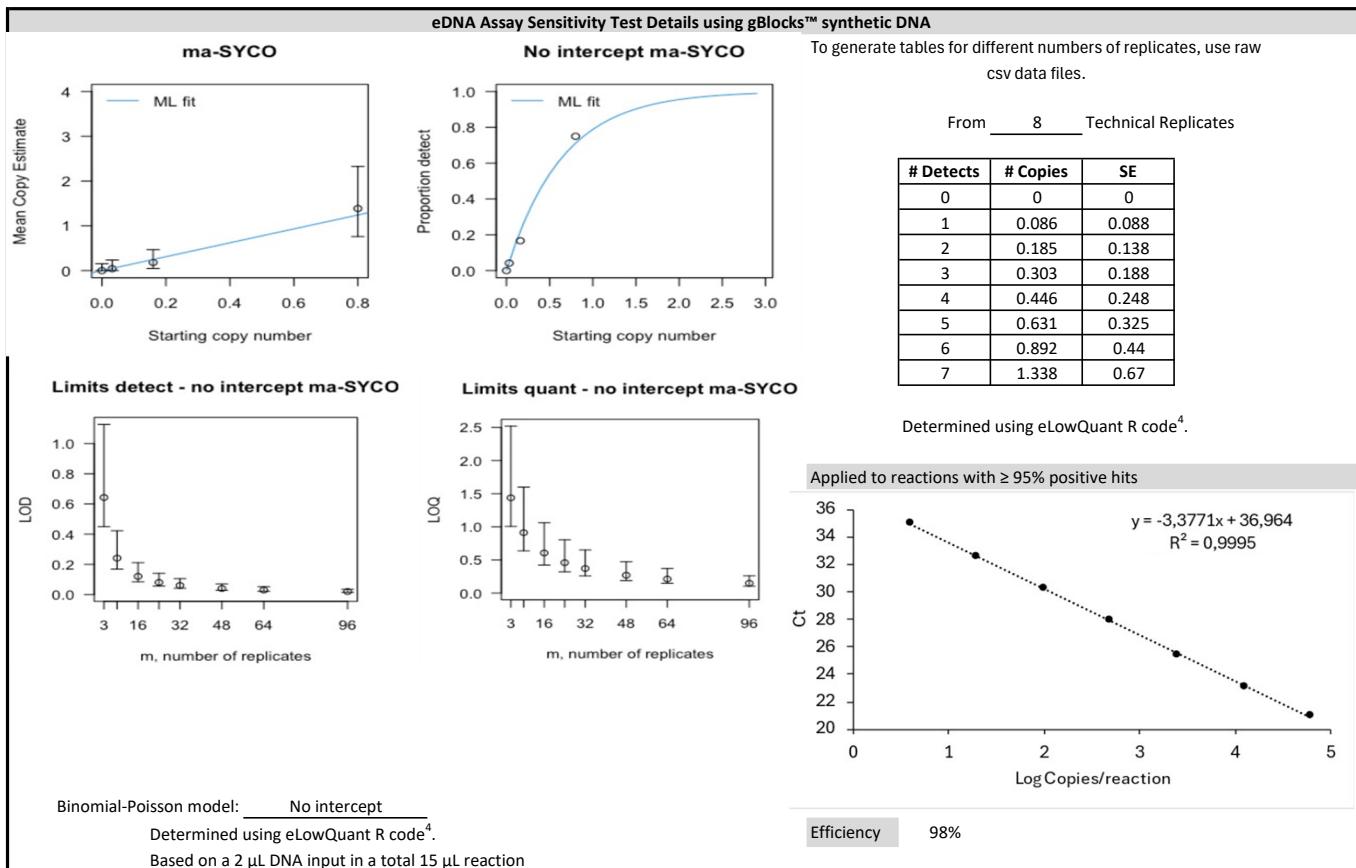
Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

##### # Voucher

Species	Common Name (Species)	Detection	Specimens	Sample Sources/Locations
ma-SYCO	Southern bog lemming ( <i>Synaptomys cooperi</i> )	Yes	2	Quebec
ma-MIPI	Woodland vole ( <i>Microtus pinetorum</i> )	No	2	Quebec
ma-ZAHU	Meadow jumping mouse ( <i>Zapus hudsonius</i> )	No	3	Quebec
ma-SYBO	Northern bog lemming ( <i>Synaptomys borealis</i> )	No	1	Quebec
ma-CLGA	Gapper's red-backed vole ( <i>Clethrionomys gapperi</i> )	No	3	Quebec
ma-PEsp	Deer mouse or White-footed mouse ( <i>Peromyscus sp.</i> )	No	3	Quebec
ma-NAIN	Woodland jumping mouse ( <i>Napoeozapus insignis</i> )	No	3	Quebec
ma-HOSA	Human ( <i>Homo sapiens</i> )	No	1	Cell line ATCC
ma-CALUfa	Domestic dog ( <i>Canis lupus familiaris</i> )	No	1	Quebec
ma-FECA	Domestic cat ( <i>Felis catus</i> )	No	2	Quebec

#### **References**

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA, 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 3: 970-981. doi: 10.1002/edn3.220



Field Sample Validation					
Known					
Sample Type	Presence	# Samples	Detected	Location	

Abbreviations					
95% CI	95% Confidence interval		LOQ	Limit of quantification	
eDNA	Environmental DNA		MT-ND2	Mitochondrial NADH dehydrogenase 2	
gDNA	Total genomic DNA extracted from voucher specimen		NTC	qPCR no template control	
LOB	Limit of blank		qPCR	Quantitative real-time polymerase chain reaction	
LOD	Limit of detection		SE	Standard error	