



## Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity<sup>1-3</sup>.

### General eDNA Assay Information

Target Species: Wood turtle (*Glyptemys insculpta*)  
Species Code: re-GLIN

eDNA qPCR Tool: eGLIN1  
eDNA qPCR Format: TaqMan

Gene Target: MT-CYB  
Published in:

### eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD 0.4      95% CI 0.3-0.7 Copies/Rxn      LOQ 1.7      95% CI 1.2-2.7 Copies/Rxn      LOB 0 hits/8  
LOQ<sub>continuous</sub> 4 Copies/Rxn

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code<sup>4</sup>. When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

### eDNA Assay Specificity Test Information

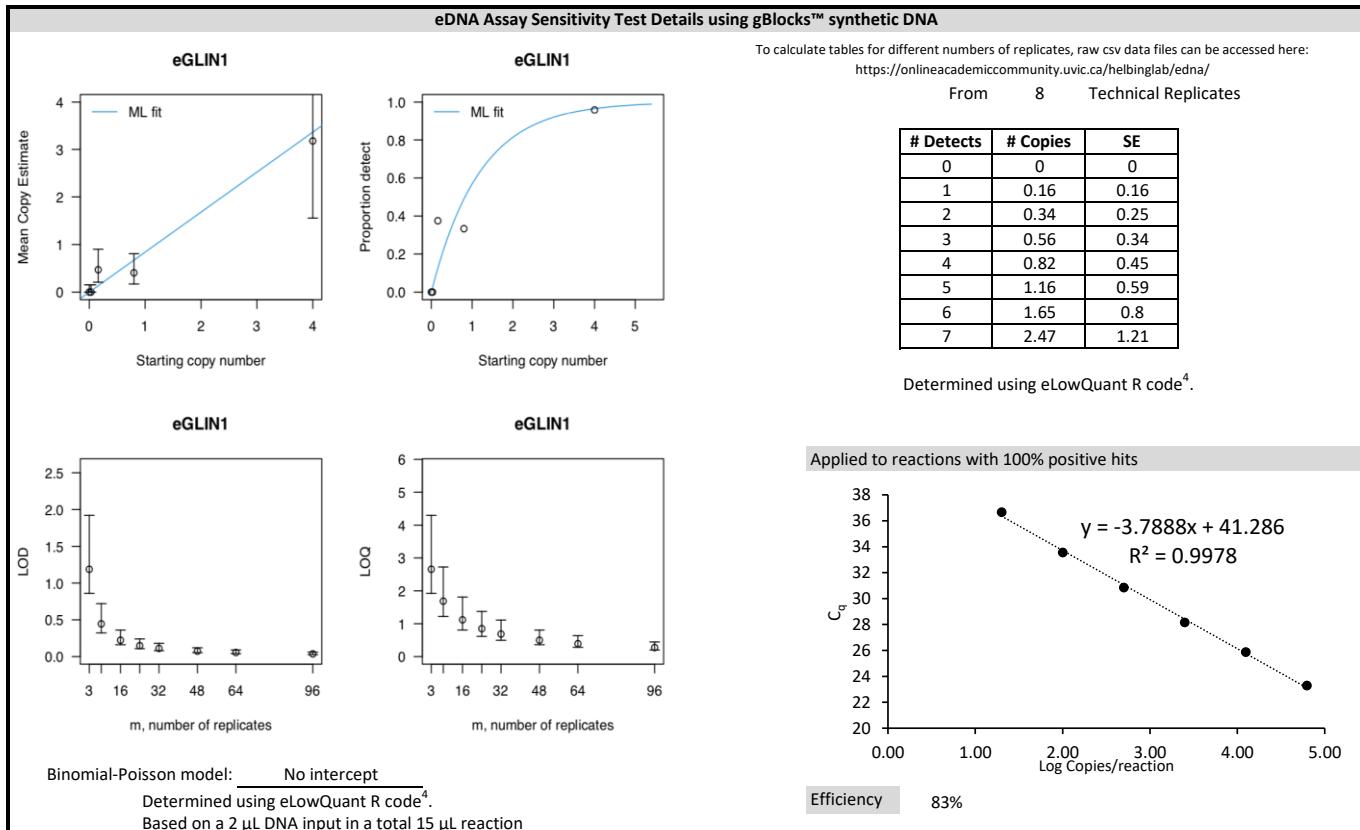
Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

#### # Voucher

Species	Common Name (Species)	Detection	Specimens	Sample Sources/Locations
am-LICA	North American Bullfrog ( <i>Lithobates (Rana) catesbeianus</i> )	No	1	British Columbia
ma-HOSA	Human ( <i>Homo sapiens</i> )	No	1	Netherlands
re-CHPI	Painted turtle ( <i>Chrysemys picta</i> )	No	1	Ontario
re-CHSE	Snapping turtle ( <i>Chelydra serpentina</i> )	No	1	Nova Scotia
re-COTE	Sharp-Tailed Snake ( <i>Contia tenuis</i> )	No	1	British Columbia
re-ELCO	Common Wall Lizard ( <i>Podarcis muralis</i> )	No	1	British Columbia
re-EMBL	Blandings turtle ( <i>Emydoidea blandingii</i> )	No	1	Nova Scotia
re-GLIN	Wood turtle ( <i>Glyptemys insculpta</i> )	Yes	2	Nova Scotia
re-POMU	Common Wall Lizard ( <i>Podarcis muralis</i> )	No	1	British Columbia
re-THEL	Western Terrestrial Garter Snake ( <i>Thamnophis elegans</i> )	No	1	British Columbia
re-THOR	Northwestern Garter Snake ( <i>Thamnophis ordinoides</i> )	No	1	British Columbia
re-THSI	Common Garter Snake ( <i>Thamnophis sirtalis</i> )	No	1	British Columbia

### References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA. 2020; 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 00: 1-12. doi: 10.1002/edn3.220



Field Sample Validation				
Known				
Sample Type	Presence	# Samples	Detected	Location
Water	Y	1	Y	Nova Scotia

Abbreviations				
95% CI	95% Confidence interval		LOQ	Limit of quantification
eDNA	Environmental DNA		MT-CYB	Mitochondrial cytochrome B gene
gDNA	Total genomic DNA extracted from voucher specimen		NTC	qPCR no template control
LOB	Limit of blank		qPCR	Quantitative real-time polymerase chain reaction
LOD	Limit of detection		SE	Standard error