



Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity¹⁻³.

General eDNA Assay Information

Target Species: Great white shark (*Carcharodon carcharias*)
Species Code: ch-CACA

eDNA qPCR Tool: echCACA2
eDNA qPCR Format: TaqMan

Gene Target: MT-ND5
Published in:

eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD	0.2	95% CI	0.1-0.3	Copies/Rxn	LOQ	0.7	95% CI	0.3-1.2	Copies/Rxn	LOB	0	hits/8
				LOQ _{continuous}	4							

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code⁴. When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

eDNA Assay Specificity Test Information

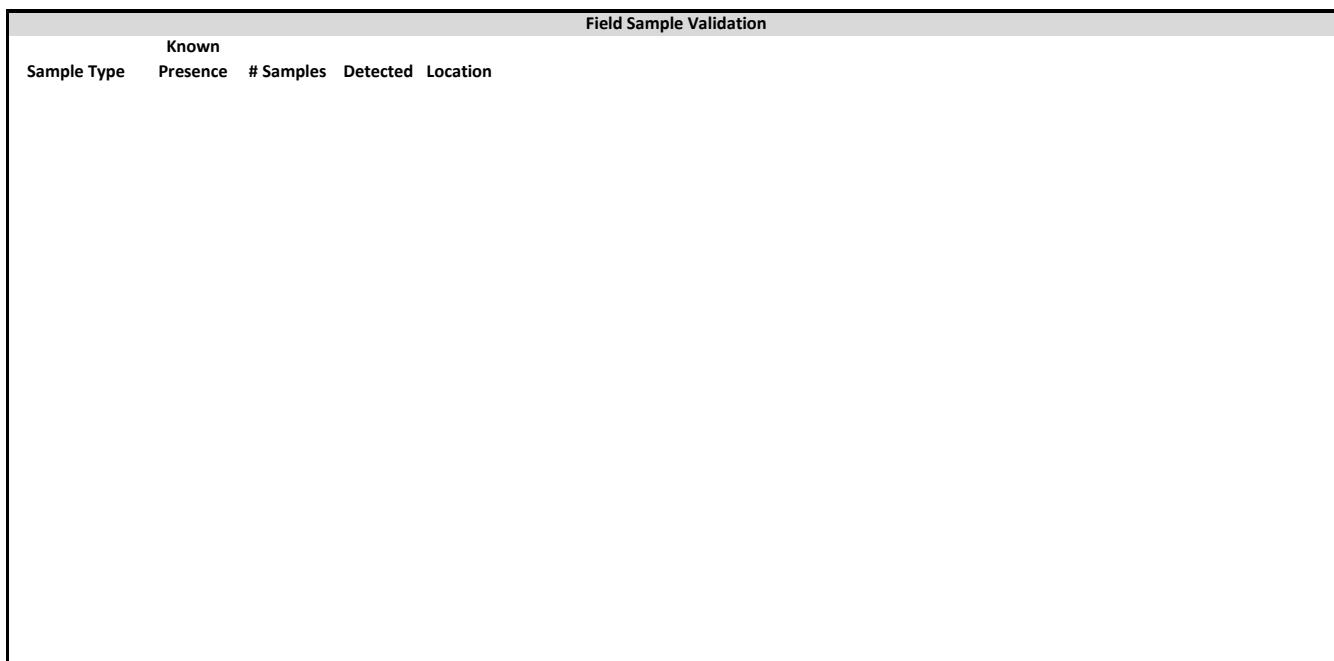
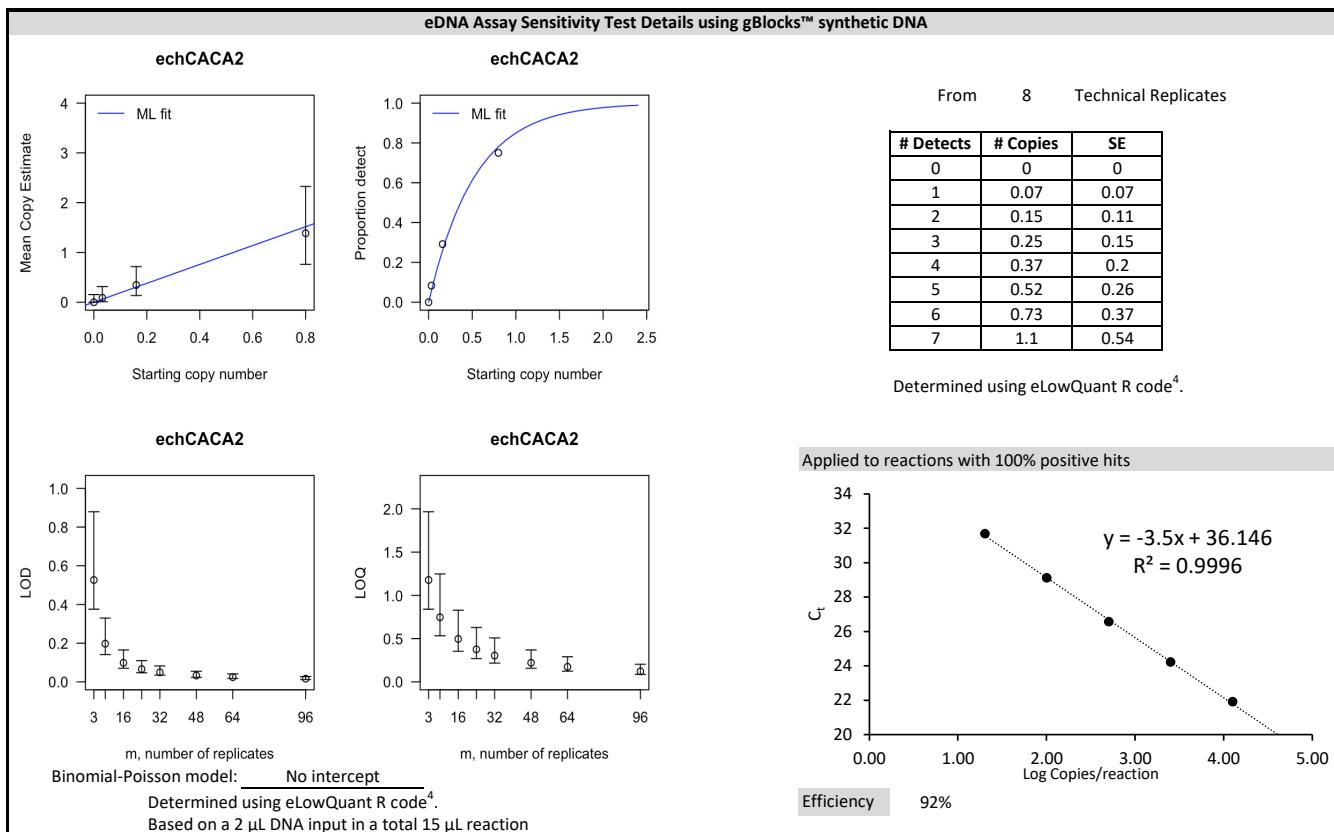
Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Voucher

Species	Common Name (Species)	Detection	Specimens	Sample Sources/Locations
ch-CACA	Great white shark (<i>Carcharodon carcharias</i>)	Yes	6	California
ch-GAGA	School/soupfin shark (<i>Galeorhinus galeus</i>)	No	1	Unknown
ch-ISOX	Shortfin mako shark (<i>Isurus oxyrinchus</i>)	No	1	Unknown
ch-LADI	Salmon shark (<i>Lamna ditropis</i>)	No	2	Unknown
ch-PRGL	Blue shark (<i>Prionace glauca</i>)	No	4	Unknown
ot-HEGR	Bluntnose sixgill shark (<i>Hexanchus griseus</i>)	No	2	Unknown
ma-CALUfa	Canine (<i>Canis lupus familiaris</i>)	No	1	British Columbia
ma-FECA	Cat (<i>Felis catus</i>)	No	1	British Columbia
ma-HOSA	Human (<i>Homo sapiens</i>)	No	1	Netherlands
te-HIST	Pacific halibut (<i>Hippoglossus stenolepis</i>)	No	1	British Columbia
te-CLPA	Pacific herring (<i>Clupea pallasi</i>)	No	1	British Columbia
te-SERU	Yelloweye rockfish (<i>Sebastodes ruberrimus</i>)	No	1	British Columbia

References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA, 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 3: 970-981. doi: 10.1002/edn3.220



Abbreviations					
95% CI	95% Confidence interval		LOQ	Limit of quantification	
eDNA	Environmental DNA		MT-ND5	Mitochondrial NADH dehydrogenase 5 gene	
gDNA	Total genomic DNA extracted from voucher specimen		NTC	qPCR no template control	
LOB	Limit of blank		qPCR	Quantitative real-time polymerase chain reaction	
LOD	Limit of detection		SE	Standard error	