



### Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity<sup>1-3</sup>.

#### General eDNA Assay Information

Target Species: *Prionospio (Prionospio (Minuspio) lighti)*  
Species Code: an-PRLI

eDNA qPCR Tool: ePRLI4  
eDNA qPCR Format: TaqMan

Gene Target: MT-CYB  
Published in:

#### eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD	0.3	95% CI	0.2-0.6	Copies	LOQ	1.2	95% CI	0.8-2.2	Copies	LOB	0	hits/8
				LOQ <sub>continuous</sub>	4				Copies/Rxn			

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code<sup>4</sup>: When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

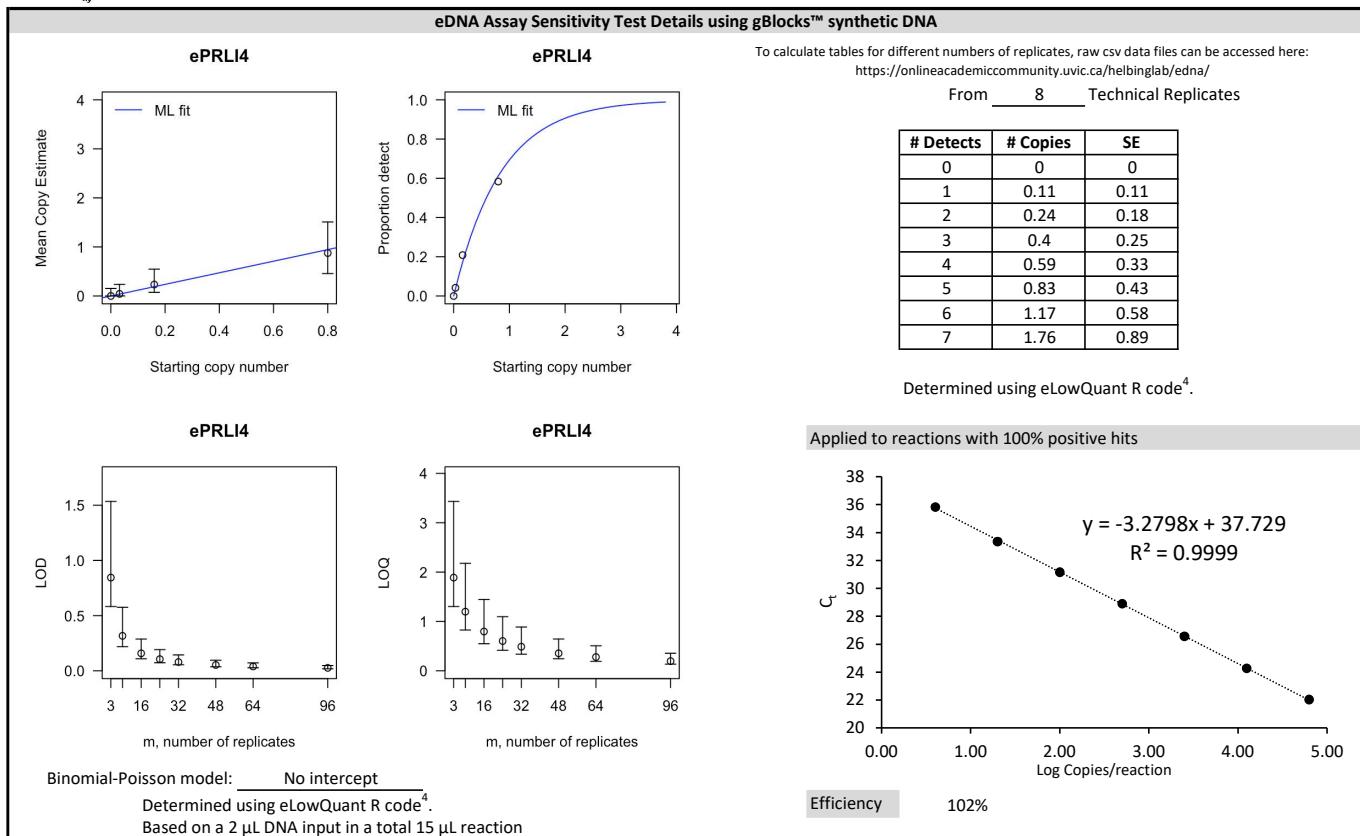
#### eDNA Assay Specificity Test Information

Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Species	Common Name ( <i>Species</i> )	# Voucher		
		Detection	Specimens	Sample Sources/Locations
an-CACAW	<i>Capitella (Capitella capitata)</i>	No	6	British Columbia
an-DEGR	<i>Decamastus (Decamastus gracilis)</i>	No	5	British Columbia
an-DRLO	<i>Drilonereis (Drilonereis longa)</i>	No	2	British Columbia
an-GLNA	<i>Glycera (Glycera nana)</i>	No	6	British Columbia
an-HEFI	<i>Heteromastus (Heteromastus filobranchus)</i>	No	6	British Columbia
an-MASA	<i>Maldane (Maldane sarsi)</i>	No	6	British Columbia
an-NOHE	<i>Notomastus (Notomastus hemipodus)</i>	No	5	British Columbia
an-OPAC	<i>Ophelina (Ophelina acuminata)</i>	No	6	British Columbia
an-PRGR	<i>Praxillela (Praxillela gracilis)</i>	No	5	British Columbia
an-PRJU	<i>Prionospio (Prionospio (Prionospio) jubata)</i>	No	6	British Columbia
an-PRLI	<i>Prionospio (Prionospio (Minuspio) lighti)</i>	Yes	6	British Columbia
an-PRMU	<i>Prionospio (Prionospio (Minuspio) multibranchiata)</i>	No	1	British Columbia
an-PRPA	<i>Praxillela (Praxillela pacifica)</i>	No	6	British Columbia
an-RIFI	<i>Hydrothermal vent worm (<i>Ridgeia picescae</i>)</i>	No	4	British Columbia
ma-CALUfa	Domestic dog ( <i>Canis lupus familiaris</i> )	No	1	British Columbia
ma-FECA	Domestic cat ( <i>Felis catus</i> )	No	1	British Columbia
ma-HOSA	Human ( <i>Homo sapiens</i> )	No	1	Netherlands

#### References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA. 2020; 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 3: 970-981. doi: 10.1002/edn3.220



Field Sample Validation				
Known				
Sample Type	Presence	# Samples	Detected	Location

Abbreviations				
95% CI	95% Confidence interval		LOQ	Limit of quantification
eDNA	Environmental DNA		MT-CYB	Mitochondrial cytochrome-B gene
gDNA	Total genomic DNA extracted from voucher specimen		NTC	qPCR no template control
LOB	Limit of blank		qPCR	Quantitative real-time polymerase chain reaction
LOD	Limit of detection		SE	Standard error