



Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity.¹⁻³

General eDNA Assay Information

Target Species: Cutthroat Trout (*Oncorhynchus clarkii*)
Species Code: te-ONCL

eDNA qPCR Tool: eONCL4
eDNA qPCR Format: TaqMan

Gene Target: MT-ND1
Published in:

eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD	0.2	95% CI	0.1-0.4	Copies/Rxn	LOQ	0.8	95% CI	0.6-1.4	Copies/Rxn	LOB	0	hits/8
				LOQ _{continuous}	4				Copies/Rxn			

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code⁴. When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

eDNA Assay Specificity Test Information

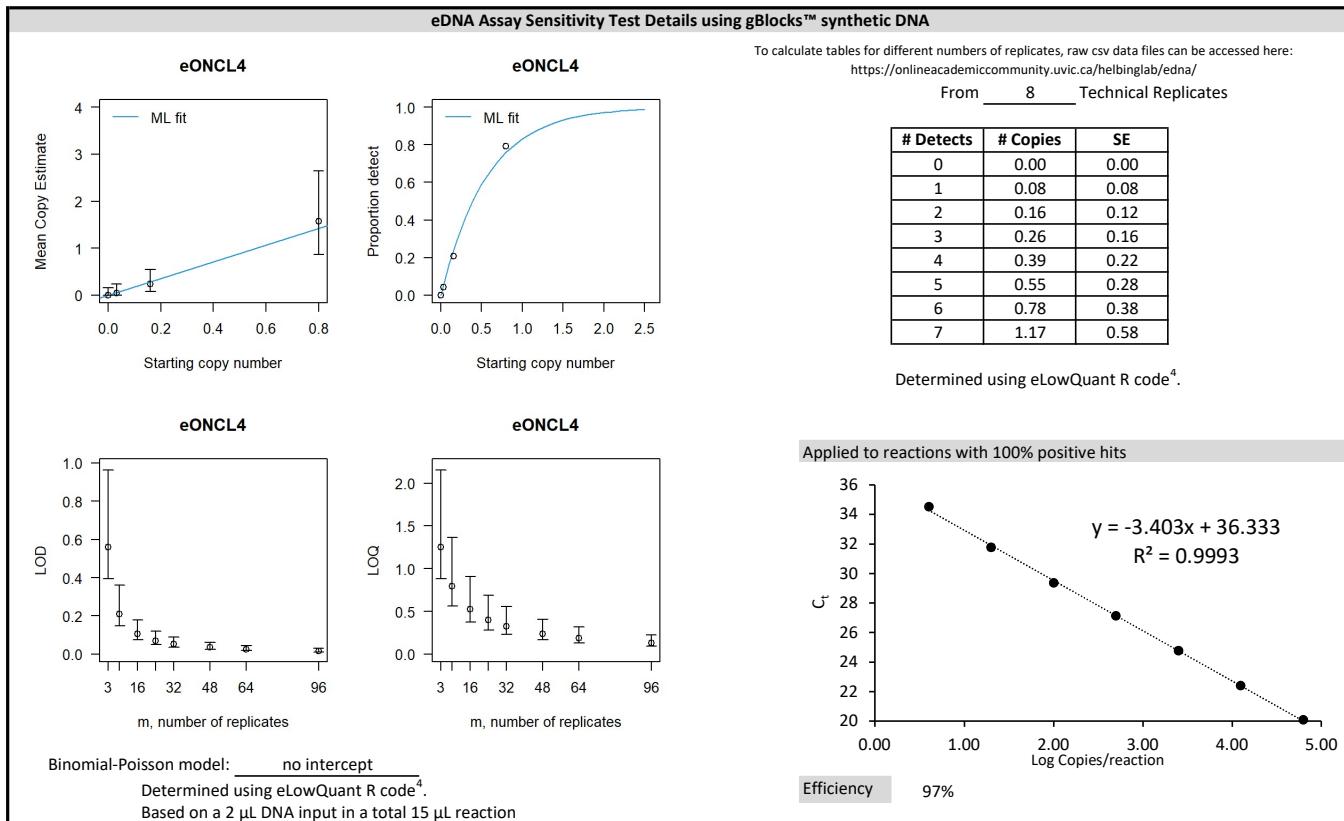
Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Voucher

Species	Common Name (Species)	Detection	Specimens	Sample Sources/Locations
ma-HOSA	Human (<i>Homo Sapiens</i>)	No	1	Netherlands
te-ONCLcl	Coastal Cutthroat Trout (<i>Oncorhynchus clarkii clarkii</i>)	Yes	5	British Columbia
te-ONCLle	Westslope Cutthroat Trout (<i>Oncorhynchus clarkii lewisi</i>)	Yes	9	Alberta
te-ONGO	Pink Salmon (<i>Oncorhynchus gorbuscha</i>)	No	1	British Columbia
te-ONKE	Chum Salmon (<i>Oncorhynchus keta</i>)	No	1	British Columbia
te-ONKI	Coho Salmon (<i>Oncorhynchus kisutch</i>)	No	1	British Columbia
te-ONMY	Rainbow Trout (<i>Oncorhynchus mykiss</i>)	No	6	Alberta and British Columbia
te-ONNE	Sockeye Salmon (<i>Oncorhynchus nerka</i>)	No	1	British Columbia
te-ONTS	Chinook Salmon (<i>Oncorhynchus tshawytscha</i>)	No	1	British Columbia
te-SACO	Bull Trout (<i>Salvelinus confluentus</i>)	No	4	Alberta
te-SAFO	Brook Trout (<i>Salvelinus fontinalis</i>)	No	4	Alberta
te-SAMA	Dolly Varden (<i>Salvelinus malma</i>)	No	1	Alberta
te-SASA	Atlantic Salmon (<i>Salmo salar</i>)	No	1	Nova Scotia

References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA. 2020; 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 00: 1-12. doi: 10.1002/edn3.220



Field Sample Validation					
Sample Type	Known				
	Presence	# Samples	Detected	Location	
95% CI	95% Confidence interval				
eDNA	Environmental DNA				
gDNA	Total genomic DNA extracted from voucher specimen				
LOB	Limit of blank				
LOD	Limit of detection				

Abbreviations					
95% CI	95% Confidence interval		LOQ	Limit of quantification	
eDNA	Environmental DNA		MT-ND1	Mitochondrial NADH subunit 1 gene	
gDNA	Total genomic DNA extracted from voucher specimen		NTC	qPCR no template control	
LOB	Limit of blank		qPCR	Quantitative real-time polymerase chain reaction	
LOD	Limit of detection		SE	Standard error	