

Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity¹⁻³.

General eDNA Assay Information

Target Species: North American Bullfrog (*Lithobates [Rana] catesbeiana*) eDNA qPCR Tool: eLICA5 Gene Target: MT-ND5
Species Code: am-LICA eDNA qPCR Format: TaqMan Published in:

eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD 0.7 95% CI 0.5-1.2 Copies LOQ 2.8 95% CI 2-4.5 Copies LOB 0 hits/8
LOQ_{continuous} 20 Copies/Rxn

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code⁴. When the LOQ < LOD, use the LOD for the LOQ. Enzyme: Immolase

eDNA Assay Specificity Test Information

Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

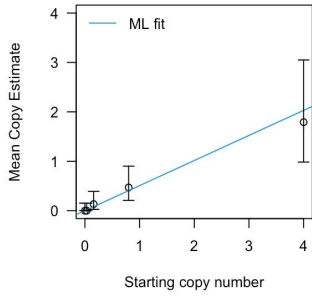
| Species | Common Name (Species) | Detection | # Voucher | |
|-----------|--|-----------|-----------|--------------------------|
| | | | Specimens | Sample Sources/Locations |
| am-LICA | Bullfrog (<i>Lithobates [Rana] catesbeiana</i>) | Yes | 5 | British Columbia |
| am-ANBO | Western Toad (<i>Anaxyrus [Bufo] boreas</i>) | No | 2 | Northwest Territories |
| am-LICL | Green frog (<i>Lithobates [Rana] clamitans</i>) | No | 1 | British Columbia |
| am-LIPI | Northern leopard frog (<i>Lithobates [Rana] pipiens</i>) | No | 2 | Alberta |
| am-LISY | Wood frog (<i>Lithobates [Rana] sylvaticus</i>) | No | 1 | British Columbia |
| am-PSRE | Pacific chorus frog (<i>Pseudacris [Hyla] regilla</i>) | No | 1 | British Columbia |
| am-PSMA | Boreal chorus frog (<i>Pseudacris maculata</i>) | No | 1 | Ontario |
| am-RAAU | Northern red-legged frog (<i>Rana aurora</i>) | No | 1 | British Columbia |
| am-RACA | Cascades frog (<i>Rana cascadae</i>) | No | 1 | British Columbia |
| am-RALU | Columbia spotted frog (<i>Rana luteiventris</i>) | No | 1 | British Columbia |
| am-RAPR | Oregon spotted frog (<i>Rana pretiosa</i>) | No | 1 | British Columbia |
| am-SPIN | Great basin spadefoot toad (<i>Spea [Scaphiopus] intermontana</i>) | No | 1 | British Columbia |
| am-XELA | African clawed frog (<i>Xenopus laevis</i>) | No | 1 | South Africa |
| am-AMGR | Northwestern salamander (<i>Ambystoma gracile</i>) | No | 1 | British Columbia |
| am-AMMA | Long toed salamander (<i>Ambystoma macrodactylum</i>) | No | 1 | British Columbia |
| am-AMMV | Eastern tiger salamander (<i>Ambystoma mavortium</i>) | No | 1 | British Columbia |
| am-ANBO | Western toad (<i>Anaxyrus [Bufo] boreas</i>) | No | 1 | British Columbia |
| am-ANAM | American toad (<i>Anaxyrus americanus</i>) | No | 1 | British Columbia |
| ma-CALUfa | Canine (<i>Canis lupus familiaris</i>) | No | 1 | British Columbia |
| ma-FECA | Cat (<i>Felis catus</i>) | No | 1 | British Columbia |
| ma-HOSA | Human (<i>Homo sapiens</i>) | No | 1 | Netherlands |

References

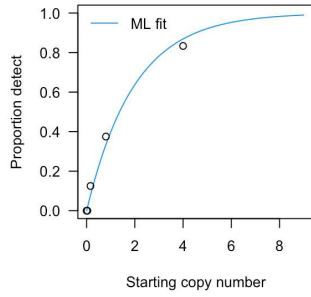
- Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA, 2: 350-361. <https://doi.org/10.1002/edn3.82>
- Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
- Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
- Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 3: 970-981. doi: 10.1002/edn3.220

eDNA Assay Sensitivity Test Details using gBlocks™ synthetic DNA

eLICA5



eLICA5

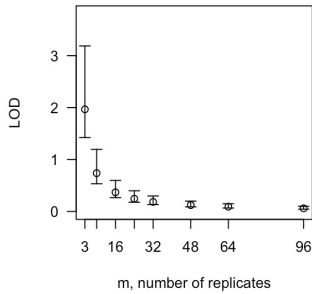


From 8 Technical Replicates

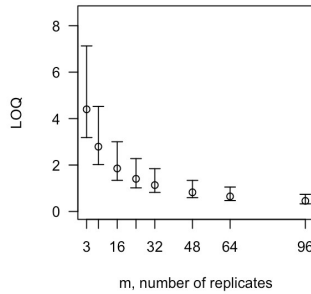
| # Detects | # Copies | SE |
|-----------|----------|------|
| 0 | 0 | 0 |
| 1 | 0.26 | 0.27 |
| 2 | 0.57 | 0.42 |
| 3 | 0.93 | 0.57 |
| 4 | 1.37 | 0.75 |
| 5 | 1.93 | 0.98 |
| 6 | 2.73 | 1.32 |
| 7 | 4.10 | 2.01 |

Determined using eLowQuant R code⁴.

eLICA5

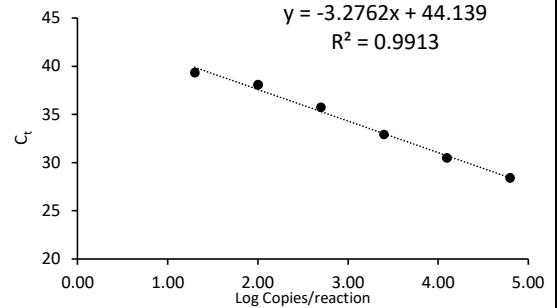


eLICA5



Binomial-Poisson model: No intercept
Determined using eLowQuant R code⁴.
Based on a 2 µL DNA input in a total 15 µL reaction

Applied to reactions with 100% positive hits



Efficiency 102%

Field Sample Validation

| Sample Type | Known | | Detected | Location |
|-------------|----------|-----------|----------|--------------|
| | Presence | # Samples | | |
| Freshwater | Y | 1 | Y | Victoria, BC |

Abbreviations

| | | | |
|--------|---|--------|--|
| 95% CI | 95% Confidence interval | LOQ | Limit of quantification |
| eDNA | Environmental DNA | MT-ND5 | Mitochondrial NADH dehydrogenase subunit 5 gene |
| gDNA | Total genomic DNA extracted from voucher specimen | NTC | qPCR no template control |
| LOB | Limit of blank | qPCR | Quantitative real-time polymerase chain reaction |
| LOD | Limit of detection | SE | Standard error |