



## Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity<sup>1-3</sup>.

## General eDNA Assay Information

Target Species: Glycera (Glycera nana)  
Species Code: an-GLNAeDNA qPCR Tool: eGLNA3  
eDNA qPCR Format: TaqManGene Target: MT-ND6  
Published in:

## eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD 0.2      95% CI 0.2-0.4      Copies/Rxn      LOQ 0.9      95% CI 0.6-1.6      Copies/Rxn      LOB 0      hits/8  
LOQ\_continuous 4      Copies/RxnBinomial-Poisson model for 8 technical replicates determined using eLowQuant R code<sup>4</sup>. When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

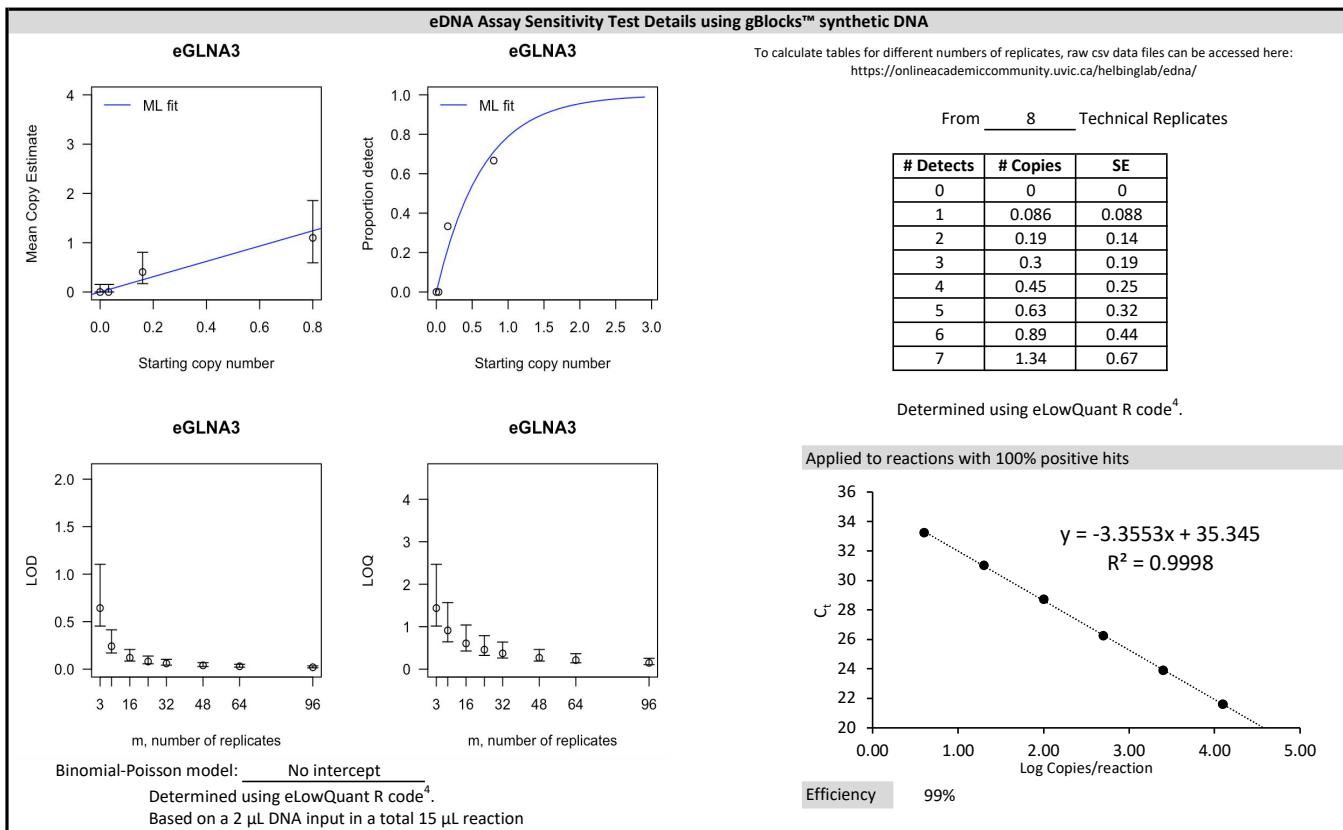
## eDNA Assay Specificity Test Information

Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Species	Common Name (Species)	Detection	# Voucher Specimens	Sample Sources/Locations
an-CACAW	Capitella (Capitella capitata)	No	2	Southwestern British Columbia
an-DEGR	Decamastus (Decamastus gracilis)	No	2	Southwestern British Columbia
an-GLNA	Glycera (Glycera nana)	Yes	5	Southwestern British Columbia
an-HEFI	Heteromastus (Heteromastus filobranchus)	No	2	Southwestern British Columbia
an-MASA	Maldane (Maldane sarsi)	No	2	Southwestern British Columbia
an-NOHE	Notomastus (Notomastus hemipodus)	No	2	Southwestern British Columbia
an-OPAC	Ophelina (Ophelina acuminata)	No	2	Southwestern British Columbia
an-PRJU	Prionospio (Prionospio jubata)	No	2	Southwestern British Columbia
an-PRLI	Prionospio [Prionospio (Minuspilo) lighti]	No	2	Southwestern British Columbia
an-PRPA	Praxillela (Praxillela pacifica)	No	1	Southwestern British Columbia
an-RUPI	Hydrothermal vent worm ( <i>Ridgeia piscesae</i> )	No	2	Southwestern British Columbia
ma-CAFA	Canine ( <i>Canis lupus familiaris</i> )	No	1	Southwestern British Columbia
ma-FECA	Cat ( <i>Felis catus</i> )	No	1	Southwestern British Columbia
ma-HOSA	Human ( <i>Homo sapiens</i> )	No	1	Netherlands
si-THNI	Segmented marine worm ( <i>Thysanocardia nigra</i> )	No	2	Southwestern British Columbia

## References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA, 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 3: 970-981. doi: 10.1002/edn3.220



## Field Sample Validation

Sample Type	Known	Presence	# Samples	Detected	Location
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Abbreviations			
95% CI	95% Confidence interval	LOQ	Limit of quantification
eDNA	Environmental DNA	MT-ND6	Mitochondrial NADH dehydrogenase subunit 6 gene
gDNA	Total genomic DNA extracted from voucher specimen	NTC	qPCR no template control
LOB	Limit of blank	qPCR	Quantitative real-time polymerase chain reaction
LOD	Limit of detection	SE	Standard error