



Helbing Laboratory eDNA Technical Bulletin

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity¹⁻³.

General eDNA Assay Information

Target Species: Zebra mussel (*Dreissena polymorpha*) eDNA qPCR Tool: eDRPOS Gene Target: MT-TG (tRNA Gly)
Species Code: mo-DRPO eDNA qPCR Format: TaqMan Published in:

eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA

LOD	0.3	95% CI	0.2-0.5	Copies/Rxn	LOQ	1.1	95% CI	0.8-2	Copies/Rxn	LOB	0	hits/8
				LOQ_continuous	4				Copies/Rxn			

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code⁴. When the LOQ < LOD, use the LOD for the LOQ.

Enzyme: QIAcuity

eDNA Assay Specificity Test Information

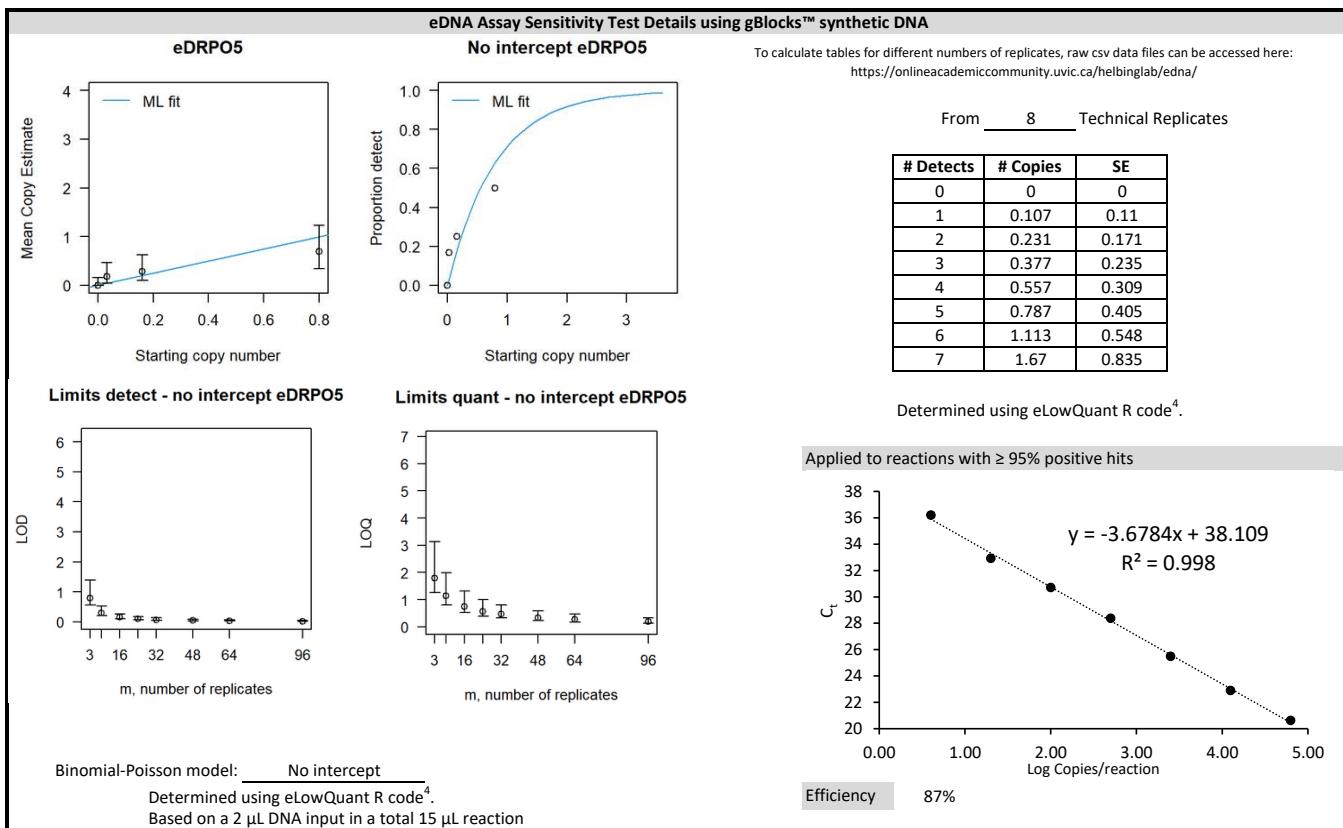
Each qPCR reaction in the specificity assay contained 10 picograms of voucher target gDNA (n=25 technical replicates)

Voucher

Species	Common Name (Species)	Detection	Specimens	Sample Sources/Locations
ma-CALUfa	Dog (<i>Canis lupus familiaris</i>)	No	1	British Columbia
ma-FECA	Cat (domestic) (<i>Felis catus</i>)	No	1	British Columbia
ma-HOSA	Human (<i>Homo sapiens</i>)	No	1	Netherlands
mo-COFL	Asian clam (<i>Corbicula fluminea</i>)	No	1	British Columbia
mo-CRGI	Pacific oyster (<i>Crassostrea gigas</i>)	No	1	British Columbia
mo-DRBU	Quagga mussel (<i>Dreissena bugensis</i>)	No	4	Ontario
mo-DRPO	Zebra mussel (<i>Dreissena polymorpha</i>)	Yes	4	Ontario
mo-MAIN	Pointed Macoma (<i>Macoma Iniquinita</i>)	No	1	British Columbia
mo-MYAR	Softshell clam (<i>Mya Arenaria</i>)	No	1	British Columbia
mo-Myspp	Mussel (<i>Mytilus spp.</i>)	No	1	British Columbia
mo-NUOB	Mahogany clam (<i>Nuttallia obscurata</i>)	No	1	British Columbia
mo-OBOL	Freshwater mussel (<i>Obiovaria olivaria</i>)	No	1	Quebec
mo-OSLU	Olympia oyster (<i>Ostrea lurida</i>)	No	1	British Columbia
mo-VEPH	Manila clam/Japanese littleneck (<i>Venerupis philippinarum</i>)	No	1	British Columbia
te-PEOM	Trout-perch (<i>Percopsis omiscomaycus</i>)	No	1	Ontario
mo-ANCA	California floater (<i>Anodonta californiensis</i>)	No	1	British Columbia

References

1. Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA, 2: 350-361. <https://doi.org/10.1002/edn3.82>
2. Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
3. Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
4. Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 3: 970-981. doi: 10.1002/edn3.220



Field Sample Validation					
Sample Type	Known	Presence	# Samples	Detected	Location
Water	Y	Y	6	N	Manitoba
Water	Y	Y	12	Y	Manitoba

Abbreviations					
95% CI	95% Confidence interval		LOQ	Limit of quantification	
eDNA	Environmental DNA		MT-TG	Mitochondrial tRNA glycine gene (tRNA Gly)	
gDNA	Total genomic DNA extracted from voucher specimen		NTC	qPCR no template control	
LOB	Limit of blank		qPCR	Quantitative real-time polymerase chain reaction	
LOD	Limit of detection		SE	Standard error	