



**Helbing Laboratory eDNA Technical Bulletin**

All eDNA tools are validated through a rigorous multi-step evaluation protocol that includes tests of DNA target specificity and amplification sensitivity<sup>1-3</sup>.

**General eDNA Assay Information**

Target Species: Lake whitefish (*Coregonus clupeaformis*) eDNA qPCR Tool: eCOCL1 Gene Target: MT-ND2  
Species Code: te-COCL eDNA qPCR Format: TaqMan Published in: \_\_\_\_\_

**eDNA Assay Sensitivity Test Summary using gBlocks™ Synthetic DNA**

LOD 0.6 95% CI 0.4-1 Copies/Rxn LOQ 2.2 95% CI 1.6-3.7 Copies/Rxn LOB 0 hits/8  
LOQ<sub>continuous</sub> 20 Copies/Rxn

Binomial-Poisson model for 8 technical replicates determined using eLowQuant R code<sup>4</sup>. When the LOQ < LOD, use the LOD for the LOQ. Enzyme: Immolase

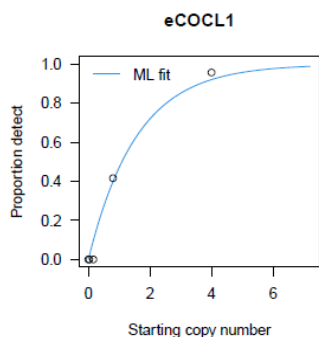
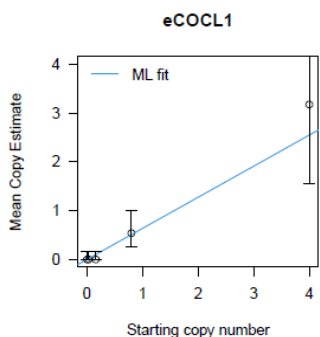
**eDNA Assay Specificity Test Information**

Species	Common Name ( <i>Species</i> )	Detection	# Voucher		Sample Sources/Locations
			Specimens		
ma-CALUfa	Dog ( <i>Canis lupus familiaris</i> )	No	1		British Columbia
ma-HOSA	Human ( <i>Homo sapiens</i> )	No	1		Netherlands
te-CACA	Longnose sucker ( <i>Catostomus catostomus</i> )	No	2		British Columbia
te-CACO	White sucker ( <i>Catostomus commersonii</i> )	No	2		Alberta
te-CAMA	Largescale sucker ( <i>Catostomus macrocheilus</i> )	No	2		British Columbia
te-COCL	Lake whitefish ( <i>Coregonus clupeaformis</i> )	Yes	1		Alberta
te-MIDO	Smallmouth bass ( <i>Micropterus dolomieu</i> )	No	2		British Columbia
te-MISA	Largemouth bass ( <i>Micropterus salmoides</i> )	No	2		British Columbia
te-ONCLle	Westslope cutthroat trout ( <i>Oncorhynchus clarkii lewisi</i> )	No	2		Alberta
te-ONGO	Pink salmon ( <i>Oncorhynchus gorbuscha</i> )	No	2		British Columbia
te-ONKE	Chum salmon ( <i>Oncorhynchus keta</i> )	No	1		British Columbia
te-ONKI	Coho salmon ( <i>Oncorhynchus kisutch</i> )	No	1		British Columbia
te-ONMY	Rainbow trout ( <i>Oncorhynchus mykiss</i> )	No	2		British Columbia
te-ONNE	Sockeye salmon ( <i>Oncorhynchus nerka</i> )	No	1		British Columbia
te-PRCY	Round whitefish ( <i>Prosopium cylindraceum</i> )	No	1		British Columbia
te-SACO	Bull trout ( <i>Salvelinus confluentus</i> )	No	2		British Columbia
te-SAFO	Brook trout ( <i>Salvelinus fontinalis</i> )	No	2		British Columbia
te-SANA	Lake trout ( <i>Salvelinus namaycush</i> )	No	2		British Columbia
te-SAVI	Walleye ( <i>Sander vitreus</i> )	No	1		Alberta
te-THAR	Arctic grayling ( <i>Thymallus arcticus</i> )	No	2		Alberta

**References**

- Hobbs, J, Adams, IT, Round, JM, Goldberg, CS, Allison, MJ, Bergman, LC, Mirabzadeh, A, Allen, H, Helbing, CC (2020) Revising the range of Rocky Mountain tailed frog, *Ascaphus montanus*, in British Columbia, Canada, using environmental DNA methods. Environmental DNA. 2020; 2: 350-361. <https://doi.org/10.1002/edn3.82>
- Hobbs, J, Round, JM, Allison, MJ, Helbing, CC (2019) Expansion of the known distribution of the coastal tailed frog, *Ascaphus truei*, in British Columbia, Canada, using robust eDNA detection methods. PLOS ONE 14(3): e0213849. <https://doi.org/10.1371/journal.pone.0213849>
- Langlois, VS, Allison, MJ, Bergman, LC, To, TA, and Helbing, CC (2020) The need for robust qPCR-based eDNA detection assays in environmental monitoring and risk assessments. Environmental DNA, 3: 519-527. doi: 10.1002/edn3.164
- Lesperance, M, Allison, MJ, Bergman, LC, Hocking, MD, and Helbing, CC (2021) A statistical model for calibration and computation of detection and quantification limits for low copy number environmental DNA samples. Environmental DNA, 00: 1-12. doi: 10.1002/edn3.220

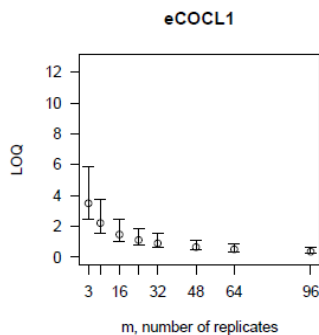
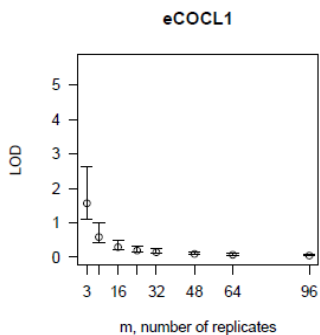
**eDNA Assay Sensitivity Test Details using gBlocks™ synthetic DNA**



From 8 Technical Replicates

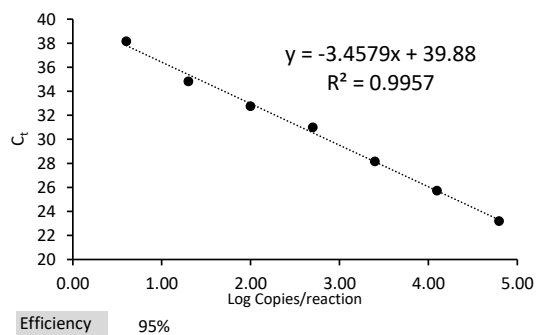
# Detects	# Copies	SE
0	0	0
1	0.21	0.21
2	0.45	0.33
3	0.74	0.46
4	1.1	0.6
5	1.54	0.78
6	2.17	1.06
7	3.26	1.61

Determined using eLowQuant R code<sup>4</sup>.



Binomial-Poisson model: No intercept  
 Determined using eLowQuant R code<sup>4</sup>.  
 Based on a 2 µL DNA input in a total 15 µL reaction

Applied to reactions with 100% positive hits



**Field Sample Validation**

Sample Type	Known Presence	# Samples	Detected	Location
-------------	----------------	-----------	----------	----------

**Abbreviations**

95% CI	95% Confidence interval	LOQ	Limit of quantification
eDNA	Environmental DNA	MT-ND2	Mitochondrial NADH dehydrogenase subunit 2 gene
gDNA	Total genomic DNA extracted from voucher specimen	NTC	qPCR no template control
LOB	Limit of blank	qPCR	Quantitative real-time polymerase chain reaction
LOD	Limit of detection	SE	Standard error